

1 **Foetal Allogeneic Intracerebroventricular Neural Stem Cell Transplantation in**
2 **People with Secondary Progressive Multiple Sclerosis: A phase I dose-**
3 **escalation clinical trial**

4 Leone MA^{1*}, Gelati M^{1*}, Profico DC¹, Conti C², Spera C², Muzi G², Grespi V¹, Bicchi I¹, Ricciolini C¹, Ferrari D⁴,
5 Zarrelli M¹, Amoruso L¹, Placentino G¹, Crociani P¹, Apollo F¹, Di Viesti P¹, Fogli D¹, Popolizio T¹, Colosimo C²,
6 Frondizi D², Stipa G², Tinella E², Ciampini A², Sabatini S², Paci F², Silveri G¹, Gobbi C^{3,6}, Pravatà E^{3,6,7}, Zecca E^{3,6},
7 Balzano RF⁷, Kuhle J⁵, Copetti M¹, Fontana A¹, Carella M¹, D'Aloisio G¹, Abate L¹, Ventura Carmenate Y⁸,
8 Pluchino S⁹, Peruzzotti-Jametti L⁹ and Vescovi AL^{1,4}

9 * The two Authors contributed equally to the manuscript

10

- 11 1. IRCCS Casa Sollievo della Sofferenza, Viale Cappuccini 1, 71013 San Giovanni Rotondo, Foggia, Italy
12 2. AOSP Santa Maria, via Tristano di Joannuccio 1, 05100, Terni, Italy
13 3. Multiple Sclerosis Centre (MSC), Department of Neurology, Neurocentre of Southern Switzerland, EOC,
14 6900 Lugano, Switzerland
15 4. BTBS Università Bicocca, piazza della Scienza 2, 20126, Milan, Italy
16 5. Department of Neurology, University Hospital Basel, and University of Basel/Switzerland
17 6. Faculty of biomedical Sciences, Università della Svizzera Italiana (USI), 6900 Lugano, Switzerland
18 7. Department of Neuroradiology, Neurocentre of Southern Switzerland, EOC, 6900 Lugano, Switzerland
19 8. Abu Dhabi Stem Cell Center, Abu Dhabi, United Arab Emirates
20 9. Department of Clinical Neurosciences and NIHR Biomedical Research Centre, University of Cambridge,
21 CB2 0QQ Cambridge, United Kingdom.

22

23 Corresponding Author: Prof. Angelo Luigi Vescovi. Email vescovia@gmail.com

24 IRCCS Casa Sollievo della Sofferenza, Viale Cappuccini 1, 71013 San Giovanni Rotondo, Foggia, Italy Tel
25 +39 0882 416566

26

27 **ABSTRACT**

28 **Background:** Advanced cell therapeutics are emerging as potentially effective treatments for chronic
29 neurological diseases, including secondary progressive multiple sclerosis (SPMS). Here we report the
30 results of a phase I trial in which good manufacturing practice-grade foetal allogeneic human neural
31 stem cells (hNSCs) were implanted via intracerebroventricular (ICV) injection in 15 individuals with
32 active and non-active SPMS.

33 **Methods:** This is a phase I, open-label, multicentre, dose-escalation, international study. The primary
34 objective was to assess the feasibility, safety, and tolerability of ICV injections of allogeneic hNSCs in
35 patients affected by SPMS over a study follow up of 12 months. We also evaluated the number and
36 type of adverse events (AEs) leading to a maximum tolerated dose, the general health status, and
37 mortality. The secondary objectives were the therapeutic benefit of allogeneic hNSCs using
38 assessment scales, magnetic resonance imaging (MRI), and laboratory and neurophysiologic
39 parameters.

40 **Findings:** Fifteen unrelated SPMS patients were enrolled and treated between 2018 and 2020. The
41 participants had a median age of 49.8 years. Their mean extended disability status scale (EDSS) at
42 enrolment was 7.6, the mean disease duration was 22 years, and mean time from diagnosis to
43 progression was 10.1 years. Neither treatment-related deaths nor serious AEs were reported during
44 the study (1 year follow up after treatment). All the other AEs were classified as non-serious and
45 were associated to non-study concomitant therapy or other medical conditions not connected to the
46 experimental treatment. During the study, none of the participants worsened in the progression of
47 their SPMS as shown by the evaluation scales implemented to assess their progress. Laboratory and
48 neurophysiologic parameters showed no clinically significant variations. MRI follow-up showed non-
49 clinically significant type 1, 2, and 3 changes.

50 **Interpretation:** The intracerebroventricular injection of foetal allogeneic hNSCs in people with SPMS
51 is feasible, tolerated and safe. Study participants displayed a substantial clinical stability during the
52 12-month follow-up. The absence of relevant adverse reactions (Ars) arising from the
53 transplantation of hNSCs indicates a short-term neutral balance between benefits and risks and
54 suggests a concrete, though perspective therapeutic possibility for SPMS patients. Further studies
55 are needed to confirm and extend the findings herein and evaluate the actual therapeutic potential
56 of advanced cell therapeutics for a condition where the lack of effective disease modifying therapies
57 is a major unmet clinical need.

58

59 INTRODUCTION

60 Multiple sclerosis (MS) is the most common cause of non-traumatic disability in young adults, with
61 over 2.5 million sufferers worldwide.¹ The last decade has seen a fast development of effective
62 disease modifying treatments (DMTs) for patients with a relapsing remitting form of MS (RRMS).
63 However, as the disease evolves into secondary progressive MS (SPMS), DMTs have limited efficacy.
64 Thus, SPMS patients currently have a major unmet need.^{2,3} A solution to the current lack of effective
65 treatments for SPMS may come from emerging cell therapy approaches, which have shown
66 promising initial results in other central nervous system (CNS) diseases, such as amyotrophic lateral
67 sclerosis (ALS) and Parkinson's disease.⁴

68 *In vivo* studies in rodent and non-human primate models of MS have shown that human neural stem
69 cells (hNSCs)⁵⁻⁸ are an effective and safe tool to induce CNS functional recovery due to their tissue
70 specificity and multiple mechanisms of action.^{9,10} hNSCs not only repair the damaged CNS by
71 replacing cells lost to injury, but also exert immunomodulatory actions on both innate and adaptive
72 immune responses via secretion of trophic factors, cross-correction of missing enzymatic activities,
73 and metabolic reprogramming.¹¹⁻¹⁴ Additional data suggest that delivering these cells directly to the
74 CNS via a single intracerebroventricular (ICV) injection allows maximising the number of cells that
75 reach the CNS¹⁵⁻¹⁷ and may be key to target the intense compartmentalised inflammation that drives
76 progression in SPMS.¹⁸ Indeed, following ICV injection, transplanted hNSCs spread throughout the
77 ventricular and subarachnoid space,^{16,19} enabling their inflammation-guided migration into the CNS,
78 where they may reach axons and myelinating cells directly without crossing the blood brain barrier.²⁰

79 The ICV cell injection is based on a widely used, technically simple, rapid, and standardised
80 neurosurgical procedure with a minimal rate of complications (6.8%), mainly due to catheter
81 malposition, haemorrhage, and infection (Morgenstern et al., 2016) . ICV is also a relatively
82 standardised experimental procedure, since it has already been used for the injection of growth
83 factors in ALS patients²¹ and for chemotherapeutic agents in anti-tumour therapy.²²

84 Here, we report the results of the first phase I, open-label, multicentre, dose-escalation study in
85 which good manufacturing practice (GMP)-grade foetal allogeneic human neural stem cells (hNSCs)
86 were implanted via intracerebroventricular (ICV) injection in 15 individuals with SPMS. To the best of
87 our knowledge, this is also the largest phase I trial with hNSCs in people with SPMS conducted to
88 date.

89 **METHODS**

90 **Study design**

91 In this phase I, open-label, multicentre, international, dose-escalation study, the participating
92 patients were affected by SPMS with progressive accumulation of disability after an initial relapsing
93 course, with or without disease activity. Fifteen patients, between 18 and 60 years of age were
94 enrolled according to a “standard” dose-escalation phase I design following a modified Fibonacci
95 sequence (100%, 60% and 50% dose increments). After the initial screening, all screened patients
96 entered a 3-month run-in phase. After that, the patients were prospectively enrolled into four
97 cohorts receiving four different doses of allogeneic hNSCs (5, 10, 16 and 24 million cells).

98 This study was performed in three participating centres located in Italy and Switzerland. The Italian
99 centres, the “IRCCS Casa Sollievo della Sofferenza” Research Hospital (Site 1) and the “Santa Maria di
100 Terni” Hospital (Site 2), recruited the patients. The MSC of the Neurocentre of Southern Switzerland
101 performed the magnetic resonance imaging (MRI) analysis, while ICV treatment was performed at
102 Site 2.

103 The study was approved by the Ethical Committee of the Istituto Tumori “Giovanni Paolo II” (Bari)
104 from the “Fondazione IRCCS Casa Sollievo della Sofferenza” Research Hospital (01PU/2016–21-01-
105 2016); the Ethical Committee of the “Aziende Sanitarie dell’Umbria” (2404/17); the Agenzia Italiana
106 del Farmaco (AIFA); the Istituto Superiore di Sanità (3090(16)-PRE21-1408–06-04-2016). The trial
107 was subsequently registered in the European Clinical Trials Database (EudraCT, 2015-004855-37),
108 and in ClinicalTrials.gov (NCT03282760).

109 **Participants**

110 Eligible patients were adults of either sex with a diagnosis of SPMS, with or without disease activity
111 (Lublin 2014) with an Expanded Disability Status Score (EDSS)²³ ≥ 6.5 and ≤ 8 , showing a progressive
112 accumulation of disability after initial relapsing course over the 2 years before recruitment (≥ 1.0
113 point for patients with EDSS =6.5 at the time of inclusion, and ≥ 0.5 points for patients with EDSS $>$
114 6.5 at the time of inclusion), and ineligibility to other therapeutic alternatives (as assessed by the
115 treating neurologist). All patients signed a written informed consent to be enrolled in the study.

116 Exclusion criteria included: other neurologic conditions; psychiatric/personality disorders or severe
117 cognitive decline; history of significant systemic, infectious, oncologic, or metabolic disorders; other
118 autoimmune diseases; chronic infections (HBV, HCV, HIV, tuberculosis); inability to undergo MRI
119 scans; inability to provide informed consent, received immunomodulant/immunosuppressive
120 treatments <6 months before inclusion; participated in other research; any contra-indication to
121 lumbar puncture; and were pregnant or breast feeding.

122 At the end of the run-in period, and if no serious co-morbidity nor health status changes occurred,
123 patients were deemed eligible for the intervention. After eligibility assessment and provision of
124 informed consent, each eligible patient was registered in the “Database for Clinical Studies with
125 Gene and Somatic Therapy” of the “Istituto Superiore di Sanità” (ISS). Before registration, the ISS
126 verified patient eligibility to ensure criteria were respected. Upon registration, the database
127 assigned a unique identification number to each patient which was used for anonymisation
128 purposes.

129 To establish baseline clinical features, a 3-month run-in period was started after the screening
130 examination. All patients were evaluated at the onset and the end of the run-in period by
131 investigating and performing physical and neurological examination, vital signs, pregnancy test in
132 fertile women, haematological and urine tests, lumbar puncture for standard CSF examination and
133 JC Virus test, serum and CSF collection (stored at minus 80 degrees), motor, sensory and visual
134 evoked potentials, optical coherence tomography (OCT), EDSS, MFSC, RAO brief repeatable battery,

135 MS-QOL54 for the evaluation of quality of life (Solari et al. 1999), brain and spinal MRI. At the end of
136 the run-in period, and if no serious co-morbidity nor health status changes occurred, patients were
137 deemed eligible for the intervention. Around 50% of patients had at least one new or one enlarging
138 T2-visible lesion, while 43% of patients had at least one lesion with contrast enhancement during
139 the run-in period.

140 **Study objectives and outcomes**

141 The primary objective of the study was to assess the feasibility, safety, and tolerability of foetal
142 allogeneic hNSCs delivered via ICV injection in people with active and non-active SPMS patients by
143 evaluating the following outcomes: mortality, the number and type of adverse reactions (ARs) or
144 events (AEs) leading to a maximum tolerated dose.

145 The secondary objectives were to evaluate the functional effects of hNSC therapeutics by monitoring
146 the following outcomes of disease progression: functional disability via EDSS and Multiple Sclerosis
147 Functional Composite (MSFC)²⁴ ; annualised relapse rate and time to confirmed relapse; cognitive
148 function by Rao's brief repeatable battery [BRB] of neuropsychological tests; visual, sensory and
149 motor functions via combination of electrophysiological measurements (visual, somato-sensory,
150 motor evoked potentials [EP]); and optical coherence tomography (OCT). Brain MRI evaluations
151 were performed to monitor structural changes related to both the intervention and disease activity.
152 MRIs were acquired at the two recruiting centres, using a Philips Ingenia scanner (Philips Medical
153 Systems, Best, The Netherlands) and a Siemens Verio scanner (Siemens, Erlangen, Germany).
154 Sequences type and parameters were harmonised between the two vendors to improve
155 reproducibility. The MRIs were performed at run-in onset (month -3), run-in end (month 0), and at
156 months +1, +2, +3, +4, +5, +6, +9 and +12.

157 Potential effects of the treatment on biomarkers of neuronal loss and inflammation was also
158 evaluated. Cerebrospinal fluid (CSF) and serum neurofilament (NfL) levels were used as markers of
159 neurodegeneration/neuronal damage, while CHI3L1 (also known as YKL-40) as indicator of reactive
160 astrocytes.²⁵ We also investigated serum and CSF levels of IL-17a, IL-2, IL-8, TNF- α , CCL2, CCL3,

161 CX3CL1, VEGF-a, OPN and GFAP as additional exploratory objectives. All the biomarkers were
162 evaluated in both CSF and serum collected before and after treatment (see table 1 for time points)

163 **Clinical evaluation**

164 After ICV injection, the participants were followed up for 12 months post-treatment (**Table 1**).
165 General health status and mortality evaluations were based on the occurrence of any serious co-
166 morbidities or changes in the general health of the participants post-intervention and the number of
167 deaths due to the treatment, or the procedure itself.

168

169 Table 1: Clinical trial study plan

	Screening	Run in START	Run in END	Surgery	M1	M2	M3	M4	M5	M6	M7	M8	M9	M10	M11	M12
Informed consent signature	x	x														
Medical history	x															
Neurological history	x															
Physical examination	x	x	x													
Infectious screening	x															
Vital Signs	x	x	x		x		x			x			x			x
Pregnancy test for fertile women	x	x	x													
Hematological tests	x	x	x		x	x	x	x	x	x	x	x	x	x	x	x
Electrocardiogram	x															
Chest X-ray	x															
standard urine test	x	x	x		x		x			x			x			x
Neurological examination	x	x	x		x	x	x	x	x	x	x	x	x	x	x	x
VAS		x	x		x	x	x	x	x	x	x	x	x	x	x	x
EDSS		x	x		x		x			x			x			x
MSFC		x	x		x		x			x			x			x
Evoked potential test		x	x		x		x			x			x			x
OCT		x	x		x		x			x			x			x
Brain and spinal MRI		x	x	x	x	x	x	x	x	x			x			x
RAO battery		x	x		x		x			x			x			x
MS-QOL54		x	x		x		x			x			x			x
Conselling with psychologist	x				x											
Serum collection for biomarkers		x	x		x		x			x			x			x
CSF collection for biomarkers		x	x		x					x						x
JC virus examination		x	x		x					x						x
AE record				x		x	x	x	x	x	x	x	x	x	x	x
Thin slice cranial CT or MRI				x	x											

170

171 All AEs were recorded and evaluated for their relationship with the hNSCs or injection procedure.

172 Each AE was classified and categorised according to the International Conference on Harmonisation

173 guidance for Clinical Safety Data Management’s Definitions and Standards for Expedited Reporting

174 E2A and the Common Terminology Criteria for Adverse Events (CTCAE). Clinical relapses were

175 defined as the appearance of a new neurologic deficit, or worsening of previously stable or

176 improving pre-existing neurologic deficit, separated by at least 30 days from the onset of a preceding
177 clinical demyelinating event.²⁶ The neurologic deficit must have been present for at least 24 hours
178 and occurred in the absence of fever (<37.5°C) or known infection.

179 **hNSCs dosage and production**

180 hNSCs were produced following a previously described method²⁷ and in full compliance with the
181 conditions and practices required by GMP regulations. The hNSCs consisted of a highly enriched
182 population of cells extracted from a single female foetal human donor (spontaneous miscarriage
183 16weeks after conception) under Ethics Committee approval, with the donors' parents' informed
184 consent, and according to the Helsinki declaration. hNSCs were cultured for 10-17 passages *in*
185 *vitro*.²⁷, the last passage was performed 24-96 hours before formulation of the final drug product.

186 On the day of treatment, hNSCs were collected from culture flasks, centrifuged counted and
187 suspended in HBSS at a concentration of 50,000 cells/ μ l. After batch release, the cells were
188 maintained at 4 +/-2 °C for up to 1.5 hours prior to implantation. See Profico et al 2022 for complete
189 quality control strategy for batch release.

190 **Intervention**

191 Trial participants were admitted to the Neurosurgery Unit of the "Azienda Ospedaliera Santa Maria"
192 (Terni, Italy) for the ICV injection of hNSCs. Before the intervention, they underwent thin-slice cranial
193 CT or MRI scans to evaluate the ventricular system and plan the procedure. Image guidance was
194 provided by the frameless stereotactic AxiEM system (Stealth station AxiEM electromagnetic
195 tracking system, Medtronic navigation, Louisville, CO, USA) to perform the ventricular cannulation.
196 The correct placement of the catheter was verified based on the egress of CSF. Then, a Rickham
197 reservoir was connected to the ventricular catheter. The participants were then prospectively
198 assigned to receive 5 (n=3), 10 (n=3), 16 (n=3), and 24 (n=6) x 10⁶ cells ICV. All the participants
199 underwent a post-operative CT scan within 24 hours to evaluate any complications.

200 All the participants received methylprednisolone orally 125 mg 2 hours pre-intervention, and
201 cefazolin 1g IV immediately before and after the hNSCs injection. The immunosuppressive treatment

202 also included oral prednisone with a 28-day taper (consisting of a dose change per week, from 60,
203 40, 20, to 10 mg q.d). The participants also received Tacrolimus (0.05 mg/kg, oral, b.i.d.), 12 hours
204 after the intervention and then every 12 hours, for 6 months. This drug was titrated to maintain
205 blood levels ranging 5–10 ng/ml.

206 **Data management and study monitoring**

207 All trial demographic and clinical data was collected by designated investigators at the screening,
208 run-in, and follow-up visits using ad-hoc case report forms (CRFs). A clinical trial monitor periodically
209 reviewed the CRFs with source data verification and requested corrections as needed. The data were
210 then entered into a eCRF and underwent systematic quality control of the study manager prior to
211 database locking. The database was periodically reviewed by an independent Data and Safety
212 Monitoring Board (DSMB) to ensure protocol adherence and to monitor possible AEs. The DSMB
213 made recommendations concerning the continuation, modification, or termination of the trial as
214 needed.

215 **Statistical Methods**

216 Demographic, clinical, and laboratory patients' characteristics were reported as median and
217 interquartile ranges (IQR) or as mean and standard deviation (SD), as appropriate, for continuous
218 variables and as frequency and percentage for categorical variables. Normal distribution was
219 checked using the Shapiro-Wilk test. Safety analyses were performed in all subjects receiving at least
220 one injection of hNSCs. All AEs and severe AEs (SAEs) were recorded at follow-up visits and at the
221 end of the study. Exploratory efficacy analyses were conducted fashion in all subjects receiving at
222 least one injection of hNSCs (here, FAS and ITT populations matched). Because of small sample size
223 and of almost all variables having non-normal distributions, pre-post differences (12-month visit vs
224 run-in end visit) were assessed via linear models using ranks. New or enlarging T2-visible lesions and
225 lesions with contrast enhancement were analysed using negative binomial models with follow-up
226 time as the offset and results reported as annualised rates.

227 A p-value <0.05 was considered statistically significant. All analyses have been performed using SAS

228 Release 9.4, SAS Institute, Cary, NC, USA.

229 **RESULTS**

230

231 **Clinical Results**

232 A total of 220 candidates applied to participate in this study between September 26, 2017, and
233 January 13, 2020. After undergoing the screening visit and completing the run-in period, fifteen
234 SPMS patients were enrolled in the study. The participants had a median age of 49.8 years (range:
235 37.8–56.6), eight of them were females and seven were males (1.14 : 1 ratio), and an almost equal
236 proportion was recruited at the two study sites. The mean EDSS was 7.6 (range 7–8), mean disease
237 duration was 22 years (range 16–29), and mean time from diagnosis to progression was 10.1 years
238 (range: 1–20). All the patients' detailed demographic and clinical characteristics are reported in
239 **Table 2.**

240 Table 2: Demographic data and group assignment.

DOSE	Gender	EDSS	AGE AT TREATMENT (range)
5*10 ⁶	M	7,5	55-59
	M	7	50-54
	M	8	50-54
10*10 ⁶	M	7,5	36-39
	F	7	50-54
	F	8	50-54
16*10 ⁶	F	7	50-54
	F	8	50-54
	F	8	45-49
24*10 ⁶	F	7,5	50-54
	F	8	45-49
	M	7	40-45
	F	7	55-59
	M	8	36-39
	F	8	50-54

241 No deaths nor serious ARs were reported during the study period. The AEs that occurred, including
242 their severity, relationship to study and non-study therapies, actions taken, and outcomes are
243 reported in **Table 3**.

244 Table 3: Adverse events

Patient	Description of event	expected?	Severity	Relationship to study therapy	Action taken	If other, please specify	New treatment /therapy given/taken?	Outcome (at the end of the study)
#1	Tremor	Yes	Mild	Unrelated	Tacrolimus dose changed		No	Recovered
	Flu like syndrome	No	Mild	Unrelated	None		Yes	Recovered
	Respiratory failure	Yes	Mild	Unrelated	Concomitant therapy changed		Yes	Recovered
	Cataract (left eye; surgical procedure)	No	Mild	Unrelated	None		No	Recovered
	Upper respiratory infection	No	Mild	Unrelated	Concomitant therapy changed		Yes	Recovered
#2	Hyperglycemia	Yes	Moderate	Unrelated	Concomitant therapy changed		No	Recovered
	Urinary retention	Yes	Moderate	Unrelated	Other, specify	Catheterization	Yes	Recovering (improving)
	Back pain	No	Mild	Unrelated	Other, specify	New therapy given	Yes	Recovered
#4	Seizure	Yes	Mild	Possible	Other, specify	Diazepam, levetiracetam	Yes	Recovered
#7	Psychosis	No	Moderate	Unrelated	Concomitant therapy changed		Yes	Recovered
#8	Fracture (right femur)	No	Moderate	Unrelated	Other, specify	Follow-up 10 omitted	No	Recovered with sequelae
#9	Leukoencephalopathy	No	Mild	Unrelated	Other, specify	MRI evaluations	No	Recovering (improving)
	Urinary tract infection	No	Mild	Unrelated	Other, specify	Drugs administration	Yes	Recovered
#12	Depression	No	Moderate	Unrelated	None		No	Unknown
#14	Urinary tract infection	No	Mild	Unrelated	Other, specify	Antibiotics administration	Yes	Not recovered

245 Among these, SAEs (defined as AEs requiring hospitalisation) occurred in 2 patients (2/15), but none

246 of them were related to the hNSC injection: of these two, one patient developed a steroid-induced

247 acute psychiatric disorder 1 month post-injection, but recovered completely within 1 month with the

248 administration of valproate, lorazepam, olanzapine, and psychotherapy; Another patient
249 experienced femur fracture during a physiotherapy session at 18 months post-injection.

250 Other AEs were classified as non-serious and only one was possibly related to the study. This was a
251 patient (1/15) who experienced a first-ever partial motor seizure at month 6 (expected, possibly
252 related to MS). As per the possible complications related to the long-term immunosuppressive
253 treatment, one patient (1/15) developed a tremor during treatment with Tacrolimus which
254 disappeared with dose-adjustment, one patient (1/15) developed respiratory failure, and infections
255 were detected in three patients (3/15) (1 upper respiratory tract, 2 urinary tract). All the other
256 reported AEs were related to non-study concomitant therapy or other medical conditions not
257 related to the experimental protocol. The immunosuppressive treatment was successfully
258 completed by patients. Tacrolimus was well tolerated by all patients except for the one already
259 mentioned. In all patients, Tacrolimus blood levels were within the therapeutic target range (below
260 20 ng/mL).

261 As per the secondary objectives of the study, no changes were measured in EDSS and MSFC for the
262 whole length of the study (Table 4).

263 Table 4. EDSS and MSFC evaluation

264	hNSC		Run-in Phase	Run-in Phase	1 month	3 months	6 months	9 months	12 months	p
265	Dose		Onset (N=15)	End (N=15)	(N=15)	(N=15)	(N=15)	(N=15)	(N=15)	value
266	5	MSFC (Total								0.753
267	millions	Score)								
268		Median	-5.5	-5.5	-5.6	-5.4	-5.6	-5.6	-5.3	
269		Nobs	3	3	3	3	3	3	3	
270		EDSS								0.114
271		Median	7.5	7.5	7.5	7.5	7.5	7.5	7.0	
272		Nobs	3	3	3	3	3	3	3	
273	10	MSFC (Total								0.150
274	millions	Score)								
275		Median	-6.2	-6.0	-6.0	-6.0	-5.9	-5.9	-5.9	
276		Nobs	3	3	3	3	3	3	3	
277		EDSS								1.000
278		Median	7.5	7.5	7.5	7.5	7.5	7.5	7.5	
279		Nobs	3	3	3	3	3	3	3	
280	16	MSFC (Total								0.924
281	millions	Score)								
282		Median	-6.0	-5.7	-6.2	-6.1	-5.8	-5.8	-5.6	
283		Nobs	3	3	3	3	3	3	2	
284		EDSS								1.000
285		Median	8.0	8.0	8.0	8.0	8.0	8.0	8.0	
286		Nobs	3	3	3	3	3	3	3	
287	24	MSFC (Total								0.772
288	millions	Score)								
289		Median	-6.1	-6.1	-6.0	-6.2	-5.9	-6.1	-6.2	
290		Nobs	6	6	6	5	3	4	4	
291		EDSS								0.831
292		Median	7.8	7.8	7.8	7.8	7.8	7.8	7.8	
293		Nobs	6	6	6	6	6	6	6	
294										

295 EDSS: Expanded Disability Status Scale; MSFC: Multiple Sclerosis Functional Composite; Std.dev: Standard Deviation; IQR: Interquartile Range
 296 (i.e. first, third quartiles)

297 Test for linear trend from Run-In End to the end of the study: p-values from linear mixed-effects models

298

299 The median EDSS score did not change from the end of the run-in phase to the end of the study. Two
300 patients (2/15) had a change in the functional systems score of >1 point, both in the pyramidal area:
301 one decreased from 4.5 to 3.0 and another one increased from 0.0 to 2.0. The MSFC scores also did
302 not significantly change from the end of the run-in phase to the end of the study.

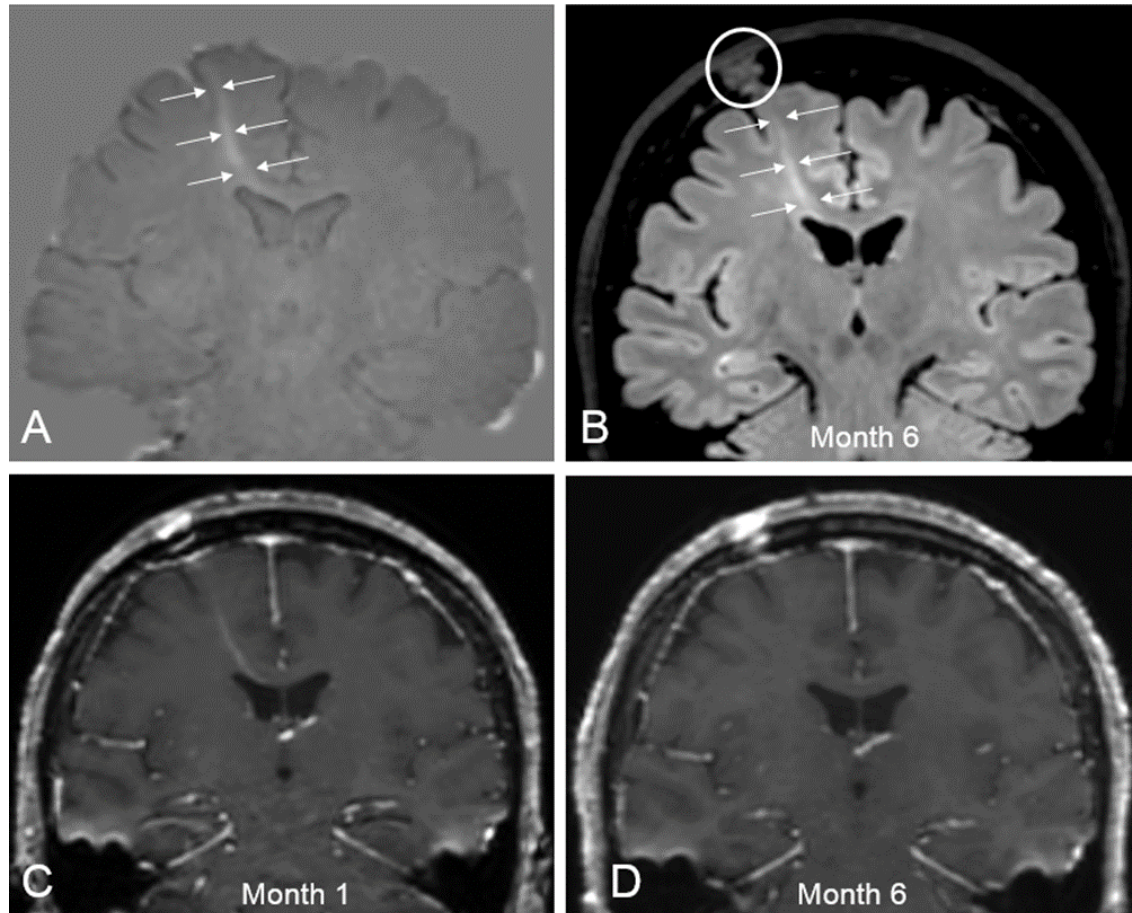
303 None of the patients reported symptoms indicative of clinically active disease and cognitive
304 functions, as measured by Rao's BRB, did not show significant changes during the study period for
305 any test. Neurophysiological parameters were monitored with EPs. Linear model analyses on ranks
306 did not show any variation trends throughout the study for any visual, somatosensory, and motor
307 EPs. We did not observe any changes on the OCT, except for one patient that showed an increase in
308 retinal nerve fibre layer's (RNFL) thickness in both eyes at month 6, which was interpreted as an
309 artefact.

310 As part of the secondary objectives, of the 40 scheduled MRIs for the 15 patients, 25 were
311 completed according to the study protocol and 15 were not performed (11 due to patient refusal,
312 two due to patient inability, two due to the COVID-19 pandemic). One patient did not have a
313 baseline MRI and post-Gadolinium images were not available for 8 MRIs due to exam interruption.

314 Brain changes on MRI were classified as type 1, 2, and 3 to facilitate comparison between time
315 points.

316 Type 1 changes were likely related to the surgical procedure. They included a linear T2-
317 hyperintensity in the parenchyma beneath the surgical right frontal cranial hole, passing through the
318 right frontal lobe white matter to the homolateral ventricle and corpus callosum (**Figure 1**).

319 **Figure 1.** Example of type 1 change, interpreted as parenchymal gliotic modifications following
320 ventricular cannulation. Coronal-oblique 3D-T2-FLAIR subtraction map in A (Month 12 minus Run-in
321 End) detected a linearly-shaped positive signal change, passing through the right frontal lobe to the
322 homolateral ventricle. This change corresponded to a subtle contrast-enhancing tract which
323 occurred at Month 1 (B, post-Gadolinium T1-weighted image) at the level of the surgical cranial hole
324 (circle in B). Month 6 (C) and Month 12 (D) T2-FLAIR images demonstrated the persisting chronic
325 changes (arrows).



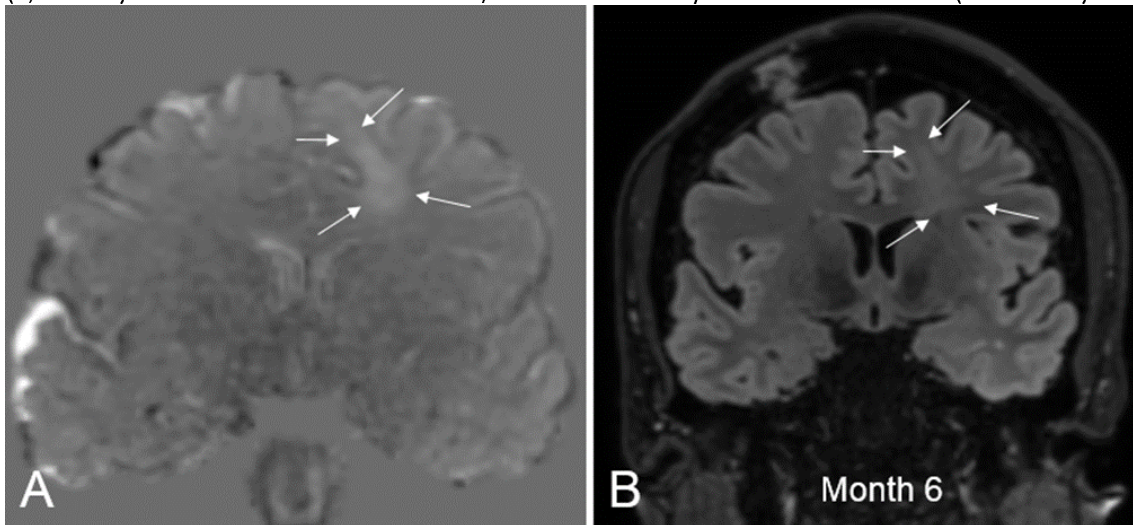
326

327 This was variable across all participants, ranging from a very subtle to a manifest signal change, and
328 it was detectable in all patients on Month 1 images, together with a linear contrast enhancement on
329 the corresponding post-Gadolinium T1-weighted images in most of cases.

330 Type 2 changes were MS-related and were used to compare the pre-transplantation with the post-
331 transplantation period.

332 Type 3 changes were undetermined and detected in only one participant, where subtraction images
333 highlighted a triangle-shaped, subtle T2-hyperintense signal change in the left frontal lobe white
334 matter, without contrast enhancement or water diffusion restriction (**Figure 2**).

335 Figure 2– Type 3 changes. Coronal-oblique 3D-T2-FLAIR subtraction map (Month 6 minus Run-in-
336 End) detects a triangle-shaped, positive signal change in the left frontal white matter (arrows). This
337 corresponded to a subtle T2-FLAIR signal hyperintensity on the corresponding Month 6 source image
338 (B, arrows). No contrast enhancement and/or water diffusivity restriction was noted (not shown).



339

340 This change remained stable at the subsequent follow-ups and was categorised as non-specific. This
341 patient had additional MRI follow-up during which the lesion did not change.

342 Pre-transplantation (during the three-month run-in period), nine new-onset and two enlarging T2-
343 visible lesions were detected in 7/14 patients (50% of patients, with an average of 1.3 and 0.3 of
344 new and enlarging T2 lesions per patient) (Table 5).

345 Table 5 Summary of pre- and post-transplantation new-onset and enlarging T2-visible lesions
 346 obtained by using subtraction imaging (see main text for details). N/A = data not available *When
 347 the Month12 time-point data was not available, the last previous available data was employed for
 348 subtraction

349

Subject #	<u>Pre-Transplantation</u>			<u>Post-Transplantation</u>		Monthly Lesion Activity Rate
	Compared Time Points	Number of New (Enlarging) T2-visible lesions	Monthly Lesion Activity Rate	Compared Time Points*	Number of New (Enlarging) T2-visible lesions	
#1	Run In End – Run In Onset	0	0.00	Month12 – Run In End	8	0.67
#2		0	0.00	Month12 - Run In End	1	0.08
#3		0	0.00	Month12 - Run In End	0	0.00
#4		2	0.67	Month12 - Run In End	6	0.50
#5		1	0.33	Month12 - Run In End	0	0.00
#6		0	0.00	Month12 - Run In End	1	0.08
#7		0	0.00	Month12 - Run In End	0	0.00
#8		0	0.00	Month 6 - Run In End	0 (1)	0.17
#9		0 (2)	0.67	Month12 - Run In End	8	0.67
#10		3	1	Month12 - Run In End	26	2.17
#11		1	0.33	Month12 - Run In End	3	0.25
#12		1	0.33	Month 3 - Run In End	0	0.00
#13		0	0.00	Month 9 - Run In End	0	0.00
#14		N/A	N/A	Month12 - Run In End	6	0.50
#15		1	0.33	Month12 - Run In End	5 (2)	0.58
Total		9 (2)			64 (3)	

350

351 After transplantation (during the 12-month follow-up period), 64 new-onset and three enlarging T2-
 352 visible lesions were detected in 10 out of 15 patients (66% of patients, with an average of 6.4 and 0.3
 353 of new and enlarging T2 lesions per patient). The annualised pre-transplantation rate of new or
 354 enlarging T2-visible lesions was 3.30 (95% CI: 1.86–5.83), which was not statistically different (p-

355 value= 0.26) to the post-transplantation rate (4.72, 95% CI: 2.32–9.61). When the 12-month data
356 was not available, the last previously available data was used to compare).
357 Pre-transplantation, 10 lesions with contrast enhancement were detected in 6/14 patients (43%)
358 (Table 6).

Subject #	Number of lesions with contrast enhancement (Time point)	Number of lesions with contrast enhancement (Time point)
#1	0	1 (Month6)
#2	0	0
#3	0	0
#4	2 (Run in Onset)	3 (Month3), 2 (Month6)
#5	1 (Run in End)	0
#6	0	0
#7	0	0
#8	0	1 (Month3)
#9	0	1 (Month6)
#10	1 (Run in Onset), 1 (Run in End)	9 (Month3), 12 (Month 6), 4 (Month12)
#11	1 (Run in End)	1 (Month9)

359
360 Table 6 Summary of pre- and post-transplantation lesions showing contrast enhancement on T1-
361 weighted images, detected upon visual inspection. N/A = data not available.

#12	1 (Run in Onset), 1 (Run in End)	0
#13	0	0
#14	N/A	1 (Month12), 1(Month6)
#15	1 (Run in Onset), 1 (Run in End)	4 (Month3)
Total	5 (Run in End), 5 (Run in Onset)	40

362

363 After transplantation, 40 gadolinium enhancing lesions were detected in multiple scans from 8 out
364 of 15 patients. The annualised pre-transplantation rate of lesions with contrast enhancement was
365 2.86 (95% CI: 1.51–5.40) and it was not statistically different (p-value=0.98) to the post-
366 transplantation values (2.83, 95% CI: 0.92–8.73).

367 Of note, 40% of the new-onset T2 lesions and the majority of gadolinium enhancing lesions
368 happened in one single patient who had the highest disease activity in the run-in period.
369 Additionally, one patient who showed one new T2 lesion and two lesions with contrast
370 enhancement in the pre-transplantation period, had no new T2 or gadolinium enhancing lesions at
371 follow-up.

372 Among the laboratory exams performed during the study, the only clinically significant variation was
373 in the number of white blood cells that ranged from a mean of $7.4 \times 10^9/L$ at the end of the run-in
374 period to a mean of $8.3 \times 10^9/L$ at the 12-month follow-up. The vital signs of the participants (blood
375 pressure and temperature) were within the normal range and urine tests did not show clinically
376 significant abnormalities. Biomarker analyses of NfL and CHI3L1/YKL-40 showed a statistically
377 significant increase in the NfL concentration (pg/mL) at 1 month after intervention in both serum
378 and CSF, followed by a gradual return to pre-intervention values (**Table 7**).

379 Table 7 Estimated marginal means, along with 95% confidence interval, from repeated measurements ANOVA models of serum neurofilament light
 380 concentrations (pg/mL) collected in NSC-SPMS patients at different groups and time points. P-values are referred to within-group and within-time
 381 comparisons and were derived from statistical contrasts defined into each model

Biomarker	hNSC assigned dose groups	Run-In onset (RSTART)	Run-In end (REND)	1 month (T1)	3 months (T3)	6 months (T6)	9 months (T9)	12 months (T12)	Within-group comparisons (p-values)						
									RSTART vs. REND	T1 vs. REND	T3 vs. REND	T6 vs. REND	T9 vs. REND	T12 vs. REND	p-value for trend
NFL CRMs	5 millions	13.84 (7.85-24.39)	11.33 (6.43-19.97)	31.61 (17.94-55.70)	29.87 (16.95-52.63)	20.62 (11.70-36.33)	12.93 (7.34-22.78)	13.21 (7.50-23.28)	0.552	0.003	0.005	0.079	0.695	0.648	0.664
	10 millions	14.59 (8.28-25.71)	16.91 (9.60-29.80)	45.64 (25.90-80.43)	21.78 (12.36-38.38)	18.21 (10.33-32.08)	17.17 (9.74-30.26)	19.68 (11.17-34.68)	0.660	0.004	0.452	0.826	0.964	0.652	0.825
	16 millions	11.80 (6.70-20.79)	12.33 (7.00-21.73)	98.03 (50.35-190.86)	27.94 (15.85-49.23)	23.07 (13.09-40.65)	14.33 (8.13-25.25)	14.84 (7.62-28.90)	0.896	<0.001	0.018	0.066	0.654	0.626	0.491
	24 millions	18.70 (12.53-27.91)	14.90 (9.98-22.24)	41.41 (27.74-61.81)	26.05 (16.91-40.12)	19.49 (11.41-33.29)	16.18 (8.51-30.79)	14.32 (8.91-23.00)	0.340	<0.001	0.030	0.370	0.814	0.882	0.540
	Overall	14.53 (11.15-18.94)	13.70 (10.51-17.86)	49.19 (37.21-65.03)	26.23 (20.06-34.29)	20.27 (15.33-26.80)	15.06 (11.23-20.20)	15.33 (11.52-20.41)	0.708	<0.001	<0.001	0.019	0.576	0.499	0.078
	Within-time comparisons (p-values)	0.594	0.743	0.090	0.881	0.948	0.905	0.778							
GFAP	5 millions	77.40 (45.16-132.65)	64.20 (37.46-110.02)	79.41 (46.33-136.08)	96.66 (56.40-165.65)	83.13 (48.51-142.47)	71.45 (41.69-122.45)	81.26 (47.41-139.26)	0.259	0.200	0.016	0.121	0.516	0.156	0.846
	10 millions	169.04 (98.64-289.70)	144.04 (84.05-246.85)	184.13 (107.44-315.55)	144.11 (84.09-246.97)	173.45 (101.21-297.26)	191.89 (111.97-328.85)	187.90 (109.64-322.01)	0.333	0.140	0.998	0.262	0.086	0.111	0.469
	16 millions	148.40 (86.59-254.32)	179.56 (104.77-307.73)	222.80 (126.50-392.41)	204.55 (119.35-350.55)	176.34 (102.89-302.20)	197.01 (114.96-337.64)	180.73 (102.61-318.32)	0.250	0.252	0.430	0.912	0.574	0.972	0.957
	24 millions	92.62 (63.28-135.57)	97.47 (66.60-142.67)	110.93 (75.79-162.36)	124.85 (84.58-184.30)	117.19 (77.00-178.37)	110.82 (70.20-174.92)	112.42 (75.25-167.96)	0.661	0.269	0.049	0.215	0.461	0.287	0.301
	Overall	115.80 (90.01-148.99)	112.79 (87.67-145.11)	137.87 (106.77-178.04)	137.34 (106.66-176.83)	131.39 (101.73-169.68)	131.53 (101.45-170.54)	132.72 (102.57-171.71)	0.732	0.015	0.014	0.061	0.070	0.051	0.056
	Within-time comparisons (p-values)	0.155	0.077	0.063	0.303	0.200	0.051	0.131							

382 The same pattern was evident across the four dose groups. These changes are likely due to post-procedure tissue damage that was detectable in all the
 383 participants on MRI imaging. On the contrary, GFAP concentration did not show any peculiar pattern and remained stable throughout the trial for all the
 384 dose groups (Table 7).

385 **DISCUSSION**

386 The ICV injection of hNSCs was safe and did not lead to clinically relevant AEs post-intervention. The
387 only SAEs that occurred were not related to hNSCs or the intervention, and no withdrawals or
388 deaths were registered. Importantly, none of the patients has shown either a clinical relapse or a
389 progression of their condition during the study period. The follow-up disability evaluation scales and
390 cognitive testing documented clinical stability of all the participants. The neurophysiologic
391 parameters and OCT also showed no significant variations. The MRI disease activity, assessed by
392 annualised rate of new or enlarging T2-visible lesions and lesions with contrast enhancement, was
393 not affected by the treatment. Finally, despite an initial fluctuation of the levels of NfL and
394 CHI3L1/YKL-40, these markers later returned to their baseline values.

395 Currently, there is a lack of treatment options for SPMS. Although another study using a similar NSC
396 preparation and method of administration is underway in another Italian centre (NCT03269071), to
397 our knowledge this manuscript represents the first report of the use of hNSCs in SPMS patients. In
398 addition, this study utilises a cell line that has been already shown safe in a prior clinical trial on ALS
399 (Mazzini et al., 2015, 2019).

400 This study has limitations inherent to its early phase, non-randomised design, and small sample.
401 Also, the fact that some MRIs could not be performed could have weakened the imaging results.
402 Nevertheless, this research provides novel information on the safety and potential effectiveness of
403 this treatment modality for SPMS. It also describes a system by which, owed to the peculiar
404 expansion technique adopted for cGMP expansion (vescovi et al, 1997), the very same hNSCs from a
405 single donor used here, can be used in a broad number of future clinical trials, thereby obviating to
406 the current outstanding issues of inter-trial variability of cell drug products.

407 It is worth noting that, unlike other clinical trials, the cells used in this study are hNSCs that
408 grow as stable, reproducible unmodified cell lines (**Table 8**)

409

410

411 Table 8: Comparison between primary culture (2014) and final product release test (2018 –
412 2020). Data for final product are expressed as mean value of the release tests conducted on
413 all the batches used for the clinical trial. See Profico et al 2022 for complete quality control
414 strategy. IP: intermediate product, FP: final product, CE: Clonal Efficiency; N: Neurons; O:
415 Oligodendrocytes; A: Astrocytes; GC: Growth Curve, expressed as the slope of the growth
416 curve..

417

	IP	FP
CE	1,70%	1,45%
N	27,90%	31,50%
A	36,10%	50,50%
O	17,70%	18,70%
GC	0,076	0,057

418 This is crucial in view of the emerging concept that one key issue in cell therapy is the inter-trial
419 standardization of the cell drug, which may be homogeneous within the same clinical protocol but
420 does vary significantly between different trials, due to the scarcity of the donor cells. The approach
421 used here provides the significant advantage that the very same cGMP hNSCs used here - with all
422 the patients receiving the same cell drug - will also be available for a number of future trials both on
423 SPMS and other disorders. To the best of our knowledge, this is the first report of such an approach
424 being possible.

425 In conclusion, NSC transplantation via ICV injection appears to be a safe procedure with neither
426 major nor short-term deleterious effects. The study participants experienced a substantial clinical
427 stability during 12 months of follow-up. The considerable absence of risks for the patients indicates a
428 short-term neutral balance between benefits and risks and a therapeutic possibility on the horizon
429 for SPMS patients. Further studies are needed to confirm and extend the findings herein and
430 evaluate the actual therapeutic potential of advanced cell therapeutics for a condition where the
431 lack of effective disease modifying therapies is a major unmet clinical need.

432 **DECLARATION OF INTERESTS**

433 All of the authors declare no conflict of interest

434 **AUTHOR CONTRIBUTIONS**

435 All the authors reviewed and approved the final manuscript

ML	Clinical coordinating investigator, Patients recruitment, data analysis, manuscript writing and approval
CS	Clinical investigator, Patients recruitment, data analysis
DCP	Clinical protocol design, regulatory affairs and administrative management of the study. Biological fluids collection and management, biomarkers analysis, manuscript writing.
MG	Clinical protocol design, regulatory affairs and administrative management of the study. Drug product release, data collection and analysis, manuscript writing.
CC, MZ, LA, GP, CP, FA, PDV, DF, GS, ET, AC, SS, FP, GS, RFB	Clinical data collection and analysis
CG, EP, EZ	Clinical protocol drafting, clinical data collection and analysis
SP, LP	Data analysis, manuscript writing
GM, VG, IB, CR, DF	Drug product production and release, data collection and evaluation Non clinical testing, manuscript drafting and revision
ALV	Study coordinator, clinical protocol concept and writing, regulatory affair, provided funding and administrative support, manuscript writing and final approval
MC, AF	Study design, data collection, statistical analysis, manuscript writing
MC	Study management, administrative and regulatory support
JK, GD, LA	Biomarker analysis, data collection and evaluation
YVC	Data analysis and results interpretation

436

437 **DATA SHARING STATEMENT**

438 All relevant data are included in this manuscript. The data that support the findings of this study are
439 available from the corresponding authors upon reasonable request.

440

441 **ACKNOWLEDGEMENTS**

442 Writing assistance in the preparation of this article was provided by Maria Carolina Rojido (Medical
443 Writing Consultant). Proofreading and submission assistance was provided by Laura C Collada Ali
444 (Medical Writing Consultant).

445

446 **FUNDING**

447 Support for the clinical study, cell therapy production writing, proofreading and submission
448 assistance was funded by IRCCS Casa Sollievo della Sofferenza Research Hospital, Italy and
449 Fondazione Revert Onlus Italy.

450 REFERENCES

- 451 1. Browne P., Chandraratna D., Angood C., et al. Atlas of Multiple Sclerosis 2013: A growing global
452 problem with widespread inequity. *Neurology*. 2014;83(11):1022-4.
- 453 2. Comi G. Disease-modifying treatments for progressive multiple sclerosis. *Mult Scler*. 2013;19(11):1428-
454 36.
- 455 3. Giovannetti A.M., Pietrolongo E., Borreani C., et al. Conversion to secondary progressive multiple
456 sclerosis: Multistakeholder experiences and needs in Italy. *PLOS ONE*. 2020;15(2):e0228587.
- 457 4. Alessandrini M., Preynat-Seauve O., De Bruin K., Pepper M.S. Stem cell therapy for neurological
458 disorders. *S Afr Med J*. 2019;109(8b):70-7.
- 459 5. Lindvall O., Kokaia Z. Stem cells in human neurodegenerative disorders--time for clinical translation? *J*
460 *Clin Invest*. 2010;120(1):29-40.
- 461 6. Vescovi A.L., Parati E.A., Gritti A., et al. Isolation and cloning of multipotential stem cells from the
462 embryonic human CNS and establishment of transplantable human neural stem cell lines by epigenetic
463 stimulation. *Exp Neurol*. 1999;156(1):71-83.
- 464 7. Ferrari D., Zalfa C., Nodari L.R., et al. Differential pathotropism of non-immortalized and immortalized
465 human neural stem cell lines in a focal demyelination model. *Cell Mol Life Sci*. 2012;69(7):1193-210.
- 466 8. Rota Nodari L., Ferrari D., Giani F., et al. Long-term survival of human neural stem cells in the ischemic
467 rat brain upon transient immunosuppression. *PLoS One*. 2010;5(11):e14035.
- 468 9. Pluchino S., Quattrini A., Brambilla E., et al. Injection of adult neurospheres induces recovery in a
469 chronic model of multiple sclerosis. *Nature*. 2003;422(6933):688-94.
- 470 10. Pluchino S., Gritti A., Blezer E., et al. Human neural stem cells ameliorate autoimmune
471 encephalomyelitis in non-human primates. *Ann Neurol*. 2009;66(3):343-54.
- 472 11. Cossetti C., Alfaro-Cervello C., Donegà M., Tyzack G., Pluchino S. New perspectives of tissue remodelling
473 with neural stem and progenitor cell-based therapies. *Cell Tissue Res*. 2012;349(1):321-9.
- 474 12. Giusto E., Donegà M., Cossetti C., Pluchino S. Neuro-immune interactions of neural stem cell
475 transplants: from animal disease models to human trials. *Exp Neurol*. 2014;260:19-32.
- 476 13. Pluchino S., Cossetti C. How stem cells speak with host immune cells in inflammatory brain diseases.
477 *Glia*. 2013;61(9):1379-401.
- 478 14. Peruzzotti-Jametti L., Bernstock J.D., Vicario N., et al. Macrophage-Derived Extracellular Succinate
479 Licenses Neural Stem Cells to Suppress Chronic Neuroinflammation. *Cell Stem Cell*. 2018;22(3):355-68.e13.
- 480 15. Zhang C., Cao J., Li X., et al. Treatment of multiple sclerosis by transplantation of neural stem cells
481 derived from induced pluripotent stem cells. *Sci China Life Sci*. 2016;59(9):950-7.
- 482 16. Takahashi Y., Tsuji O., Kumagai G., et al. Comparative study of methods for administering neural
483 stem/progenitor cells to treat spinal cord injury in mice. *Cell Transplant*. 2011;20(5):727-39.
- 484 17. Ben-Hur T., Fainstein N., Nishri Y. Cell-based reparative therapies for multiple sclerosis. *Curr Neurol*
485 *Neurosci Rep*. 2013;13(11):397.
- 486 18. Peruzzotti-Jametti L., Pluchino S. Therapy with mesenchymal stem cell transplantation in multiple
487 sclerosis ready for prime time: Commentary. *Mult Scler*. 2022;28(9):1328-9.
- 488 19. Jin K., Sun Y., Xie L., et al. Comparison of ischemia-directed migration of neural precursor cells after
489 intraatrial, intraventricular, or intravenous transplantation in the rat. *Neurobiol Dis*. 2005;18(2):366-74.
- 490 20. Morgenstern P.F., Connors S., Reiner A.S., Greenfield J.P. Image Guidance for Placement of Ommaya
491 Reservoirs: Comparison of Fluoroscopy and Frameless Stereotactic Navigation in 145 Patients. *World*
492 *Neurosurg*. 2016;93:154-8.
- 493 21. Van Damme P., Robberecht W. Developments in treatments for amyotrophic lateral sclerosis via
494 intracerebroventricular or intrathecal delivery. *Expert Opin Investig Drugs*. 2014;23(7):955-63.
- 495 22. Xu X., Chen W., Zhu W., et al. Adeno-associated virus (AAV)-based gene therapy for glioblastoma.
496 *Cancer Cell Int*. 2021;21(1):76.
- 497 23. Kurtzke J.F. Rating neurologic impairment in multiple sclerosis: an expanded disability status scale
498 (EDSS). *Neurology*. 1983;33(11):1444-52.
- 499 24. Fisher D., Beutler L.E., Williams O.B. Making assessment relevant to treatment planning: the STS
500 Clinician Rating Form. *Systemic Treatment Selection*. *J Clin Psychol*. 1999;55(7):825-42.
- 501 25. Burman J., Raininko R., Blennow K., Zetterberg H., Axelsson M., Malmeström C. YKL-40 is a CSF
502 biomarker of intrathecal inflammation in secondary progressive multiple sclerosis. *J Neuroimmunol*.
503 2016;292:52-7.

- 504 26. McDonald W.I., Compston A., Edan G., et al. Recommended diagnostic criteria for multiple sclerosis:
505 guidelines from the International Panel on the diagnosis of multiple sclerosis. *Ann Neurol.* 2001;50(1):121-7.
- 506 27. Mazzini L., Gelati M., Profico D.C., et al. Human neural stem cell transplantation in ALS: initial results
507 from a phase I trial. *J Transl Med.* 2015;13:17.
- 508 Solari, A., G. Filippini, L. Mendozzi, A. Ghezzi, S. Cifani, E. Barbieri, S. Baldini, A. Salmaggi, L. L. Mantia, M.
509 Farinotti, D. Caputo and P. Mosconi (1999). "Validation of Italian multiple sclerosis quality of life 54
510 questionnaire." *J Neurol Neurosurg Psychiatry* 67(2): 158-162.
- 511 Profico, D.C.; Gelati, M.; Ferrari, D.; Sgaravizzi, G.; Ricciolini, C.; Progetti Pensi, M.; Muzi, G.; Cajola, L.; Copetti,
512 M.; Ciusani, E.; et al. Human Neural Stem Cell-Based Drug Product: Clinical and Nonclinical Characterization.
513 *Int. J. Mol. Sci.* 2022, 23, 13425. [https:// doi.org/10.3390/ijms232113425](https://doi.org/10.3390/ijms232113425)
- 514 Mazzini L, Gelati M, Profico DC, Sorarù G, Ferrari D, Copetti M, Muzi G, Ricciolini C, Carletti S, Giorgi C, Spera C,
515 Frondizi D, Masiero S, Stecco A, Cisari C, Bersano E, De Marchi F, Sarnelli MF, Querin G, Cantello R, Petruzzelli
516 F, Maglione A, Zalfa C, Binda E, Visioli A, Trombetta D, Torres B, Bernardini L, Gaiani A, Massara M, Paolucci S,
517 Boulis NM, Vescovi AL; ALS-NSCs Trial Study Group. Results from Phase I Clinical Trial with Intraspinal Injection
518 of Neural Stem Cells in Amyotrophic Lateral Sclerosis: A Long-Term Outcome. *Stem Cells Transl Med.* 2019
519 Sep;8(9):887-897. doi: 10.1002/sctm.18-0154. Epub 2019 May 18. PMID: 31104357; PMCID: PMC6708070.